

EXHIBIT D

30 min at 37°C, either with or without preheating at 90°C for 5 min, followed by electrophoresis in a 15% denaturing gel.

For detection of *rev*-EGFP mRNA, we used a 25-mer deoxyribonucleotide probe that was complementary to the EGFP mRNA of the *rev*-EGFP fusion protein. A 29-mer deoxyribonucleotide probe was used for detection of the GAPDH transcript.

HIV-1 antiviral assay. For determination of anti-HIV-1 activity of the siRNAs, transient assays were done by cotransfection of siDNAs and infectious HIV-1 proviral DNA, pNL4-3 into 293 cells as described¹⁵. Before transfection, the cells were grown for 24 h in six-well plates in 2 ml EMEM supplemented with 10% (vol/vol) FBS and 2 mM L-glutamine, and transfected using Lipofectamine Plus reagent (Life Technologies, GibcoBRL) as described by the manufacturer. The DNA mixtures consisting of 0.5 µg siDNAs or controls, and 0.5 µg pNL4-3 were formulated into cationic lipids and applied to the cells. After one, two, three, and four days, supernatants were collected and analyzed for HIV-1 p24 antigen (Beckman Coulter, Hialeah, FL). The p24 values were calculated with the aid of the Dynatech MR5000 ELISA plate reader (Dynatech Labs Inc., Chantilly, VA). Cell viability was also assessed using a Trypan Blue dye exclusion count at four days after transfection.

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Effective expression of small interfering RNA in human cells

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In many eukaryotes, expression of nuclear-encoded mRNA can be strongly inhibited by the presence of a double-stranded RNA (dsRNA) corresponding to exon sequences in the mRNA (refs 1,2). The use of this "RNA interference" (RNAi) in mammalian studies had lagged well behind its utility in lower animals because uninterrupted RNA duplexes longer than 30 base pairs trigger generalized cellular responses through activation of dsRNA-dependent protein kinases³. Recently it was demonstrated⁴ that RNAi can be made to work in cultured human cells by introducing shorter, synthetic duplex RNAs (~20 base pairs) through liposome transfection. We have explored several strategies for expressing similar short interfering RNA (siRNA) duplexes within cells from recombinant DNA constructs, because this might allow long-term target-gene suppression in cells, and potentially in whole organisms. Effective suppression of target gene product levels is achieved by using a human U6 small nuclear RNA (snRNA) promoter to drive nuclear expression of a single RNA transcript. The siRNA-like parts of the transcript consists of a 19-base pair siRNA stem with the two strands joined by a tightly structured loop and a U₁₋₄ 3' overhang at the end of the antisense strand. The simplicity of the U6 expression cassette and its widespread transcription in human cell types suggest that this mode of siRNA delivery could be useful for suppressing expression of a wide range of genes.

The U6 snRNA promoter cassettes and si-like RNA inserts are shown in Figure 1. We previously showed that RNA expressed by RNA polymerase III from the U6+1 or U6+27 cassettes was expressed primarily as full-length transcripts and was located in the nucleus^{5,6}. U6+27 transcripts, containing the first 27 nucleotides of human U6 RNA, were capped with γ-methyl phosphates and accumulated to higher levels than U6+1 transcripts. Cassettes are designed so that short RNA coding sequences are inserted between unique *SalI* and *XbaI* sites. After the *XbaI* site, the cassette encodes a strong stem to protect the transcripts against 3'–5' exonuclease attack, then a poly(U) transcription termination sequence. However, the insertion sequences discussed later also contain their own UUUU terminator at the 3' end of the inserted sequences, terminating most transcription before the cassette-encoded stem/terminator region.

To test whether expressed si-like RNA is effective, we targeted a site in human lamin A/C mRNA that has been demonstrated to be vulnerable to synthetic siRNA⁴. The inserted sequences encoded several variants of siRNA duplexes and controls, shown in Figure 1B. Previous work on synthetic anti-lamin A/C siRNA used two independent strands with 3' unpaired tails⁴. Although it would be theoretically possible to synthesize two strands independently *in vivo*, the need to anneal the two strands could make the production

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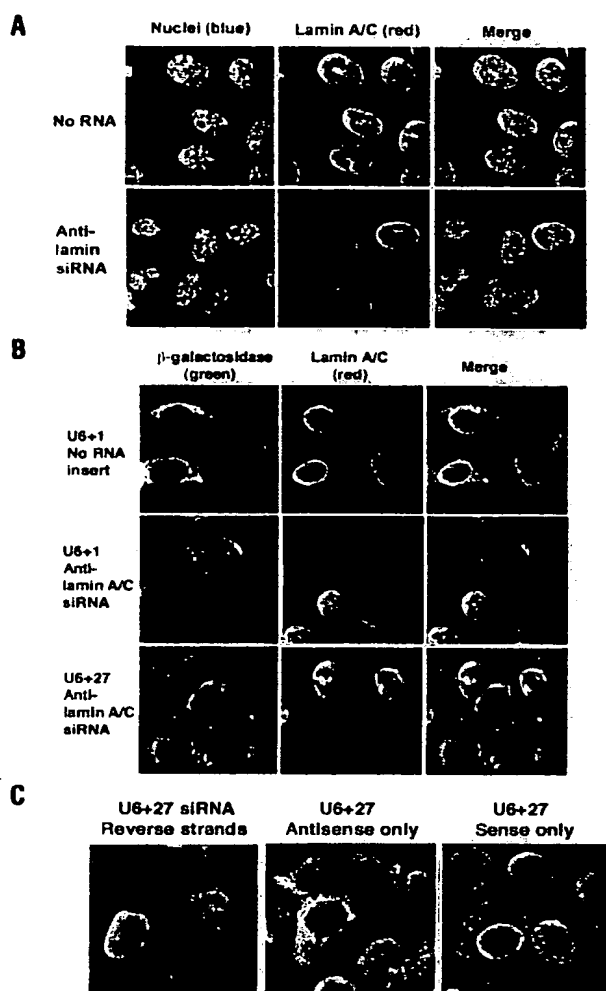




Figure 2 shows elimination of lamin A/C protein when HeLa cells were transiently transfected with either synthetic siRNA or siRNA-expressing clones. Cells shown in Figure 2A underwent Oligofectamine-mediated transfection with either no RNA or a synthetic 19-base pair siRNA duplex with 3'-TT overhangs⁴. Nuclei in Figure 2A were visualized with 4,6-diamidino-2-phenylindole (DAPI) staining (blue). As expected from previous work, the lamin A/C signal (red) substantially disappears from most cells, presumably those that are transfected. For testing recombinant DNAs, cells were cotransfected with a plasmid (pCMV β) expressing β -galactosidase (β -Gal) to mark transfected cells. Production of β -Gal also precludes the possibility that siRNA constructs nonspecifically obstruct protein synthesis. Figure 2B

Previous studies of siRNA-mediated target cleavage by extracts *in vitro* suggested that the 5' termini of one or both strands might need to be phosphorylated, and that this might be needed for efficient assembly into obligatory ribonucleoprotein complexes^{8,9}. Results pre-

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sented here suggest that the hairpin siRNAs might not need to have the 5' end of either strand unblocked. These hairpin siRNA observations would be consistent with a mechanism in which the U6 transcript containing the hairpin duplex RNA is able to assemble into any necessary protein complexes. The moderate preference for the antisense strand at the end agrees with the prediction that the antisense strand is used as a primer for RNA-dependent RNA polymerase on the message target¹⁰, but this appears not to be essential.

Another unexpected finding was that expression from the U6 snRNA promoter cassettes, which give primarily nucleoplasmic expression⁵, would succeed in inhibiting target expression when a majority of the existing mRNA is cytoplasmic. To be certain that the long hairpin did not cause altered localization, we carried out *in situ* hybridization with fluorescent probes to the hairpin that showed nuclear localization very similar to that seen previously (Fig. 3). Admittedly, a small percentage of the U6-driven tran-

Figure 3. Localization of U6+27 siRNA transcripts. Two days after transfection with the U6+27 anti-lamin A/C cassette plasmid, cells were fixed and stained for nuclear DNA (DAPI, blue) and probed with a Cy3-labeled 2'-O-methyl oligoribonucleotide (red) complementary to the antisense strand of the siRNA. As expected from work with previous U6 expression constructs^{5,6}, the U6+27 siRNA pattern was primarily in a nuclear speckled pattern. Nuclear and cytoplasmic background staining by the Cy3-oligonucleotide in the absence of U6+27-siRNA ("Mock") was minimal.

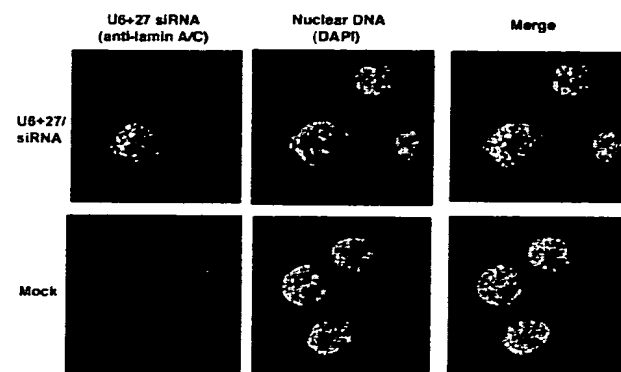
Figure 2. Effects of siRNA constructs on lamin A/C levels. HeLa cells were transfected with either synthetic siRNA or recombinant DNA cassettes expressing different small RNAs from different RNA polymerase III promoters. Cells were stained with DAPI (blue) or with antibodies to lamin A/C (red) or β -Gal (green). (A) Synthetic siRNA or no RNA transfections, showing that lamin A/C staining of the nuclear periphery is largely abolished in most cells, with only low levels of residual red staining in nuclear interiors. (B) Transfection with U6 promoter cassettes either without an siRNA insert (U6+1, no insert) or containing the anti-lamin siRNA shown in Figure 1B (U6+1 siRNA and U6+27 siRNA). Transfected cell cytoplasm is green, whereas nuclei from untransfected cells show no green cytoplasm. Empty expression cassettes have no apparent effect on lamin A/C levels (only empty U6+1 is shown), while transfected cells (green) using siRNA-expressing constructs have little remaining lamin A/C (red). (C) Overlay panels of β -Gal and lamin A/C signal after transfection with different control insertions shown in Figure 1B. Quantitative assessment of remaining lamin A/C signals in transfected cells compared to untransfected cells on the same slide is given in Table 1.

scripts might exit to the cytoplasm and be active there. These results suggest that the U6-driven transcripts are suppressing pre-mRNAs before nuclear exit.

The U6 expression cassettes used in these studies are <400 base pairs long and should be relatively easy to incorporate into a variety of vectors. The siRNA inserts can be synthesized as complementary oligodeoxynucleotide pairs to rapidly create cassettes directed at multiple sites. It should even be possible to use several cassettes per vector, targeted at either multiple mRNAs or multiple sites on the same message. It is likely that the hairpin siRNA strategy will be applicable to many mRNA targets. Preliminary experiments targeting both an endogenous human splicing factor and HIV-1 reverse transcriptase coding region (A. Ehsani, S. Li, A. Kleihauer, and J.J. Rossi, personal communication) have shown the hairpin siRNA strategy to be effective. However, as with the synthetic siRNAs, it is sometimes necessary to test several target sites along an mRNA to find one that gives the strongest inhibition. While much remains to be learned about the mechanism by which these transcripts work, the results with the simple U6 cassettes suggest that they might be useful for diverse experimental applications.

Experimental protocol

Materials. Lipofectin, Plus reagent, and Oligofectamine were purchased from Invitrogen (Carlsbad, CA), as were synthetic DNA oligonucleotides for cloning and probes. Cy3-2'-O-methyl RNA oligonucleotide hybridization probes were from Trilink (San Diego, CA). Synthetic siRNA oligonucleotides were from Dharmacon (Lafayette, CO). Anti-lamin A/C monoclonal antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA) (sc-7292, used at 1 μ g/ml); rabbit anti- β -Gal antibodies were from Molecular Probes (Eugene, OR) (A-11132, 1 μ g/ml); Oregon green 488-labeled goat anti-rabbit secondary antibodies were from Molecular Probes (O-11038, 5 μ g/ml); and cyanin-3 (Cy3)-labeled goat anti-mouse secondary antibodies were from Amersham-Pharmacia Biotech (Piscataway, NJ) (PA 43002,



1 µg/ml). Cassettes³ were cloned in pAV vectors, derived from pCWRVSVN (ref. 11) by placing the promoter modules between *Bam*HI and *Hind*III sites, after modifying the vector. Modifications included destruction of the *Bam*HI site downstream of the Neo cassette, and removal of all sites between the original *Sac*II and *Xho*I sites, inclusive, by cleavage and religation. After inserting the cassettes, a new polylinker was created between the *Hind*III and *Sac*II sites. Sequences to be expressed were inserted as synthetic oligodeoxynucleotides precisely between the end of the unique *Sac*II site and the beginning of the unique *Xba*I site. Recombinant constructs were sequenced.

Transfections. Transient transfections were carried out on subconfluent HeLa cells. Synthetic RNA was transfected using Oligofectamine as described⁴. Recombinant DNA constructs were transfected using Lipofectin with Plus reagent according to the manufacturer's instructions. In transient transfections, cells were split after one day. Cells were fixed and examined for lamin protein after three days, and fixed and examined by *in situ* hybridization after two days.

Fluorescence microscopy. Transfected cells were fixed and subjected to previously described protocols for visualizing proteins⁴ with antibodies (lamin A/C and β-Gal) or detecting small RNAs (<http://singerlab.aecom.yu.edu/protocols>) by hybridizing 5'-Cy3-labeled oligos (5'-Cy3-AAACUGGACU-UCCAGAAGAACACGAA, 2'-O-methyl ribonucleotides) to the fixed preparations. Fluorescence was acquired with a Nikon Eclipse E800 (Tokyo, Japan) with a Hamamatsu Orca II camera (Hamamatsu-City, Japan). For each construct, hundreds of cells were examined to confirm that the selected images were representative. On multiple slides, lamin A/C fluorescence in transfected cells was deconvoluted and quantitated using Isee software (Inovision; Raleigh, NC) and is expressed in Table 1 as a percentage of lamin A/C signal from nontransfected cells on the same slides. Lamin signal was consistently higher in transfected cells than in untransfected cells on the same slide.

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Using the transcriptome to annotate the genome

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A remaining challenge for the human genome project involves the identification and annotation of expressed genes. The public and private sequencing efforts have identified ~15,000 sequences that meet stringent criteria for genes, such as correspondence with known genes from humans or other species, and have made another ~10,000–20,000 gene predictions of lower confidence, supported by various types of *in silico* evidence, including homology studies, domain searches, and *ab initio* gene predictions^{1,2}. These computational methods have limitations, both because they are unable to identify a significant fraction of genes and exons and because they are unable to provide definitive evidence about whether a hypothetical gene is actually expressed^{3,4}. As the *in silico* approaches identified a smaller number of genes than anticipated^{5–9}, we wondered whether high-throughput experimental analyses could be used to provide evidence for the expression of hypothetical genes and to reveal previously undiscovered genes. We describe here the development of such a method—called long serial analysis of gene expression (LongSAGE), an adaption of the original SAGE approach¹⁰—that can be used to rapidly identify novel genes and exons.

The LongSAGE method (Fig. 1) generates 21 bp tags derived from the 3' ends of transcripts that can rapidly be analyzed and matched to genomic sequence data. The method is similar to the original SAGE approach¹⁰, but uses a different type IIS restriction endonuclease (*Mme*I) and incorporates other modifications to produce longer transcript tags. The resulting 21 bp tag consists of a constant 4 bp sequence representing the restriction site at which the transcript was cleaved, followed by a unique 17 bp sequence derived from an adjacent sequence in each transcript. Theoretical calculations show that >99.8% of 21 bp tags are expected to occur only once in genomes the size of the human genome (Table 1A). Likewise, similar analyses based on actual sequence information from ~16,000 known genes suggest that >75% of 21 bp tags would be expected to occur only once in the human genome, with the remaining tags matching duplicated genes or repeated sequences (as discussed below). In contrast, conventional SAGE tags of 14 bp do not allow unique assignment of tags to genomic sequences, though they do allow such assignment to the much less complex compendium of expressed sequence tags (ESTs) and previously characterized mRNAs^{10–12}. To optimize the quantification of transcripts, tags are ligated together to form "ditags," which are then concatenated and cloned. Sequencing tag concatemers in parallel allows the identification of up to ~30 tag sequences in each sequencing reaction. Matching tags to genome

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